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Abstract

Steroidal lactams are also a class of steroid derivatives that were synthesized and modified to increase the biological activity of steroids. Their synthesis is a stimulating challenge to the scientist, often claiming the development of new and generally useful reactions. Some of them have been tested successfully as anticancer drugs against different types of leukemia. Recent developments in the syntheses of steroidal lactams are described herein. The biological activities of those steroidal derivatives for which data are available are given.



Keywords: Steroid, oxime, lactam moiety, Beckmann rearrangement

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1. Introduction

Steroids constitute an extensive and important class of biologically active polycyclic compounds that are widely used for therapeutic purposes.¹⁻²⁷ Even after decades of research, the total synthesis of steroid nuclei by improved strategies continues to receive considerable attention. Numerous methods²⁸⁻³⁴ have been exploited for the total synthesis of steroids which are widely distributed in nature, and which possess practical medical importance. Research into steroid total synthesis continues to this day.³⁵⁻³⁸

The introduction of heteroatom or replacement of one or more carbon atoms in a steroidal molecule by a heteroatom affects the chemical properties of that particular steroid and often results in alterations of its biological activities, which sometimes may be useful. Azasteroids especially express numerous biological activities. Studies of steroidal lactams revealed that the presence of the characteristic group (-NH-CO-) in the aza-steroid molecule is important in lowering the acute toxicity and improving the anticancer activity of the compounds.³⁹

Moreover, steroidal lactams have attracted much interest, as many as 4-azalactams exhibiting strong inhibition of human steroid 5α -reductase, making them potential drugs for the treatment of benign prostatic hyperplasia (BPH), acne, male pattern baldness, and alopecia.⁴⁰ Finasteride and turosteride are two commercial drugs belonging to this class.⁴¹⁻⁴² Finasteride was the first 5α -reductase inhibitor clinically approved in 1992 in the U.S. for the treatment of BPH. The 5α -reductase inhibitory activity of these steroids is considered to be attributed to the lactam in ring A of the steroidal nucleus that mimics the intermediate transition state.⁴³

This article provides an overview of the various synthetic strategies, which have been employed to synthesize steroidal lactams along with their biological properties, if provided, from the years 2011 to 2021. Although a previous review by Panda et al. has appeared in 2013 for heterocyclic steroids, some reports were missing from their compilation regarding steroidal lactams.⁴⁴ Thus, we have chosen to cover the aforementioned period on steroidal lactams.

2. Synthesis of Steroidal Lactams

In 2011, Zhou and coworkers⁴⁵ reported the synthesis of some steroidal lactams functionalized at the 6-position of the steroidal skeleton, using cholesterol as starting material. A series of 6-substituted-3-aza-A-homo-3-oxycholestanes and 6-substituted-4-aza-A-homo-3-oxycholestanes were prepared using the Beckman rearrangement as the key step.

The oxime **4** was isolated by four steps in 66.5% yield as depicted in Scheme 1.⁴⁶ The diketone **1** was selectively reduced to yield the crude alcohol **2**, which was directly oxidized to generate 3,6-diketone **3**. In the last step,

treatment of this derivative with hydroxylamine hydrochloride (1:1.1) in ethanol in the presence of sodium acetate led to compound **4**.



Reagents : a-NaBH₄/CH₃OH, NiCl₂.6H₂O b- Jones' reagent, acetone, 88%; c- NaAc.3H₂O, 95%, ethanol, NH₂OH.HCl, 90%.

Scheme 1. Synthesis of steroidal oxime 4.

A Beckmann rearrangement of **4** with $SOCl_2/THF$ at 0 °C led to steroidal lactams **5** and **6** with moderate yields. The major product was the 4-aza derivative **5** which was isolated in 45% yield (Scheme 2).



Reagents : a-SOCI₂/THF, 0 °C, **5** (45%) and **6** (21.5%).

Scheme 2. Synthesis of steroidal lactams 5 and 6.

The steroidal lactams **7** and **11** were prepared by reduction using the reductant NaBH₄ in methanol. The oximation of compounds **5** and **6** led to steroidal derivatives **8** and **12**, respectively, as depicted in Schemes **3**

and 4. In the same way, the steroidal derivatives **9** and **10** were synthesized by treatment of compound **5** with thiosemicarbazide or semicarbazide adding a few drops of glacial acetic acid as a catalyst (Scheme 3).



Reagents : a- NaBH₄/MeOH, r.t.; b- H₂NOH.HCl/Na₂Ac.3H₂O/EtOH, reflux; c- H₂NC(S)NHNH₂/EtOH, 60 °C; d- H₂NC(O)NHNH₂/EtOH.

Scheme 3. Synthesis of steroidal lactams 7-10.



Reagents : a-NaBH₄/MeOH, r.t.; b- H₂NOH.HCl/Na₂Ac.3H₂O/EtOH, reflux.

Scheme 4. Synthesis of steroidal lactams 11 and 12.

These steroidal lactams showed distinct cytotoxicity against HeLa, MGC 7901, and SMMC 7404 cancer cells. Moreover, these results showed the importance of the functional groups in the C-6 position of the steroid skeleton for the IC50 value of these derivatives. Moreover, the authors do not observe any notable difference between the 3-aza and the 4-aza steroids. In particular, compounds **9**, **11**, **12** (IC₅₀ 6: 6.5 µmol/L; 8: 7.7 µmol/L; 9: 5.6 µmol/L) showed even higher cytotoxicity than cisplatin for HeLa cells (positive contrast, 10.1 µmol/L).

In 2012, these authors⁴⁷ reported that 6-hydroxyl-4-aza-A-homocholest-3-one **7** and 6-hydroximino-4-aza-A-homo-cholest-3-one **8**, displayed antiproliferative activity against some cancer cells through inducing cancer cell apoptosis by activation of the intrinsic pathway. Furthermore, the compound **8** was able to inhibit tumor growth in an athymic mouse model.

In 2014, the same authors⁴⁸ reported the synthesis of a series of steroidal lactams derivatives possessing different substituted groups at the C-3 position of the steroidal skeleton.

The 3-hydroxyimino steroidal derivative **18** was synthesized as depicted in Scheme 5. The synthesis of the steroid derivative **14** was already published, from compound **13**, by the same group in 2009.⁴⁹ 3-Acetoxy-7-aza-B-homocholest-4-en-6-one **15** was obtained by Beckman rearrangement of **14** with SOCl₂ in THF. Deacetylation of compound **15** in K₂CO₃ aqueous solution (13%) gave steroidal lactam **16**, which on oxidation with Jones' reagent gave 7-aza-B-homocholest-4-ene-3,6-dione **17**. Finally, the reaction of the latter with hydroxylamine hydrochloride in ethanol in the presence of NaOAc gave the steroidal lactam oxime **18** with a yield of 42%.



Reagents : a-SOCl₂/THF, 0 °C; b- K₂CO₃/MeOH, r.t., 4h; c- Jones reagent/acetone, r.t., 2h; d- NH₂OH.HCl, Na₂Ac.3H₂O, EtOH, reflux.

Scheme 5. Synthesis of steroidal lactams 15-18.

Similarly, steroidal lactams **23-26** exhibiting the 3-substituted-6-aza-B-homocholest-7-one key motif in their structures were prepared in 8 steps using cholesterol as starting material (Figure 6). The steroid derivative **19** was obtained using the procedure described in the literature.⁵⁰ The 3-keto steroid **22** was synthesized by oxidation of the alcohol **21** by Jones' reagent. Finally, the corresponding steroidal lactams **23-26** were isolated by reaction of compound **22** with HONH₂·HCl, CH₃ONH₂·HCl, PhCH₂ONH₂·HCl, or thiosemicarbazide.



Reagents : a-SOCl₂/THF, 0 °C; b- K₂CO₃/MeOH, reflux, 4h; c- Jones reagent/acetone, r.t., 2h; d- 1- NH₂OH.HCl, Na₂Ac.3H₂O, EtOH, reflux; 2- CH₃ONH₂.HCl, EtOH, 60 °C; 3- PhCH₂ONH₂.HCl, EtOH, 60 °C; 4- H₂NC(S)NHNH₂.HCl, EtOH, 60 °C.

Scheme 6. Synthesis of steroidal lactams 23-26.

Finally, the authors were interested in the synthesis of compounds **28-32** having the structure of 7 α -aza-B-homocholest-4(or 5)-ene to identify whether the aza position effects on the cytotoxicity of compounds (Scheme 7). Thus, the 3-acetoxy-7 α -aza-B-homocholest-5-ene-7-one compound **28** was obtained by Beckman rearrangement of **27**. In addition, after oxidation by Jones reagent of compound **29**, migration of its 5,6 double bond into the more stable 4,5 double bond of compound **30** is observed. Then, the reaction of the α , β -unsaturated derivative **30** with HONH₂.HCl or thiosemicarbazide led to the steroidal lactams **31** and **32** respectively.

The authors studied the antiproliferative activity of the various synthesized compounds against HeLa, SGC-7901, GNE 2, Bel-7404, Tu 686, and SPC-A cell lines. They observed remarkable cytotoxic activity for steroidal lactams possessing 3-hydroximin, 3-hydroxyl, and 3-thiosemicarbazone groups. The derivatives which have proved to be the most potent anticancer agents are the steroids **16**, **21**, **23**, **26**, and **29** which, moreover, have an antiproliferative activity similar to that of cisplatin. In addition, by the annexin V staining test, the authors showed that compound **26** was able to effectively induce tumor cell apoptosis.



Reagents : a-SOCl₂/THF, 0 °C; b- K₂CO₃/MeOH, reflux, 4h; c- Jones reagent/acetone, r.t., 2h; d- NH₂OH.HCl, Na₂Ac.3H₂O, EtOH, reflux; e- H₂NC(S)NHNH₂.HCl, EtOH, 60 °C.

Scheme 7. Synthesis of steroidal lactams 31-32.

In 2015, Cui and coworkers⁵¹ reported the synthesis of a series of 3-substituted-6-aza-B-homo-5 α -stigmastan-7-one and 3-substituted-6-aza-B-homo-5 α -sitostan-7-one derivatives using stigmasterol and sitosterol as starting materials.

Scheme 8 outlines the synthetic procedures of compounds **34a**-**42a**, **34b**-**41b** and **43b**. Compounds **33a** and **33b** were prepared as reported previously.⁵⁰ The synthesis of compounds **34a** and **34b** was carried out by Beckmann rearrangement of **33a** and **33b** with SOCl₂/THF at 0 °C, and the acetylation of compounds **34a** and **34b** gave compounds **35a** and **35b**. Next, compounds **34a**-**34b** were converted to corresponding 3-carbonyl derivatives (**36a**-**36b**) *via* oxidation with PCC. The reaction of compounds **36a** and **36b** with CH₃ONH₂.HCl, PhCH₂ONH₂.HCl or HONH₂.HCl afforded the corresponding products **37a**-**39a** and **37b**-**39b**. The (*E*)- and (*Z*)- stereoisomer were obtained in preparation of **37a**-**38a** and **37b**-**38b**, respectively. Similarly, compounds **36a** and **36b** reacted with different 4-alkyl-3-thiosemicarbazide or 4-phenyl-3-thiosemicarbazide gave the 6-aza-7-oxo-B-homo-5 α -stigmastan-3-(alkyl)thiosemicarbazone derivatives **40a**-**42a** and 6-aza-7-oxo-B-homo-5 α -sitostan-3-alkyl(or phenyl)thiosemicarbazone derivatives **40b**-**41b** and **43b**.

Their antiproliferative activities against SGC 7901 (human gastric carcinoma), CNE (nasopharyngeal carcinoma), Bel 7404 (human liver carcinoma) and HeLa (human cervical carcinoma) cancer cells were assayed. The results showed that compounds with the side chain of sitosterol had better antiproliferative activity than compounds with the side chain of stigmasterol. The compound **40b** with 3-thiosemicarbazone and **41b** with 3- (40-methyl)thiosemicarbazone group displayed an excellent antiproliferative activity against Bel-7404 cells owning an IC₅₀ value of 3.9 and 5.6 μ M, respectively (compared with cisplatin, IC₅₀: 23 μ M).







33a: ∆²², **33b**

34a: ∆²², **34b**

35a: ∆²², 35b



37a-42a ∆²² 37b-41b, 43b

37a, 37b: R = NOCH₃
38a, 38b: R = NOCH₂Ph
39a, 39b: R = NOH
40a, 40b: R = NNHCSNH₂
41a, 41b: R = NNHCSNHCH₃
42a: R = NNHCSNHCH₂CH₃
43b: R = NNHCSNHPh

Reagents : a-SOCl₂/THF, 0 °C; b- Ac₂O/Pyridine; c- PCC, CH₂C₁₂; d- NH₂OCH₃.HCl 95%, NaOAc.3H₂O, EtOH; e- PhCH₂ONH₂.HCl 95%, EtOH, NaOAc.3H₂O; f- NH₂OH.HCl 95%, NaOAc.3H₂O, EtOH; g- H₂NC(S)NHNH₂.HCl, EtOH, AcOH; h- H₂NC(S)NHNHCH₃.HCl, EtOH, AcOH; i- H₂NC(S)NHNHCH₂CH₃.HCl, EtOH, AcOH; j- H₂NC(S)NHNHPh.HCl, EtOH, AcOH.

Scheme 8. Synthesis of steroidal lactams 34-43.

In 2016, Sarli and coworkers⁵² reported the evaluation and the synthesis of novel steroidal lactam derivatives, which are conjugated with 3-(4-(bis(2-chloroethyl)amino)phenoxy)propanoic acid (POPAM) **47** hoping to improve its biological activity. Four novel steroidal lactams related to POPAM **47** have been synthesized.

The first conjugate was prepared using testosterone **44** as starting material according to the known procedure of Camoutsis and Catsoulacos (Scheme 9).⁵³ Acetylation of testosterone with acetic anhydride in pyridine led to the corresponding acetate which was condensed with hydroxylamine to provide the *Z*- and *E*-oximes **45**. In a second step, a Beckman rearrangement of the mixture of **45** with SOCl₂ in dioxane was realized and as predicted by previously described results,⁵⁴ a single lactam **46** was obtained in a moderate yield (63%) most likely due to Z/E isomerization before alkyl migration. Next, the acetyl moiety was deprotected under basic conditions with excellent yield (92%) and in a final step, the hydroxyl group at C-17 of the steroidal skeleton was esterified by POPAM **47** in the presence of DMAP and DCC in CH₂Cl₂ to provide, with a quantitative yield, the desired conjugate ENGA-L06E **48**.



testosterone ${\bf 44}$

46

45





ENGA-L06E 48





Scheme 9. Synthesis of ENGA-L06E 48.

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The second conjugate, ENGA-L08E **51** was isolated using estrone **49** as starting material as depicted in Scheme 10. The methodology of Liao and coworkers⁵⁵ was used to synthesize the 17α -aza-D-homoestrone derivative **50**. Then, esterification of lactam **50** with POPAM **47** in DMF in the presence of DCC and DMAP led to the corresponding ester **51** in good yield.



Reagents : a- 1- NH₂OH.HCl, pyridine, 100%; 2- SOCl₂, dioxane, 56%; b- POPAM **36**, DCC, DMAP, CH₂Cl₂, 68%.

Scheme 10. Synthesis of ENGA-L08E 51.

The other two steroidal lactams **57** and **61** were prepared *via* the novel monolactam **56** and dilactam **60** using adrenosterone as starting material.

The monolactam **56** was prepared as depicted in Scheme 11. Firstly, according to Morreal,⁵⁶⁻⁵⁸ a regioselective reduction of the 17-keto group of the D-ring of adrenosterone was done with NaBH₄/MeOH to provide the corresponding alcohol, which was used without further purification in the next step. Next, the hydroxyl group at C-17 was protected into its corresponding acetate **53** in a yield of 90%. The treatment of this latter with NH₂OH.HCl in pyridine led selectively to the desired oximes **54**, which by Beckman rearrangement provided the steroidal monolactam **55**. Finally, the C-17 position of compound **55** was deprotected followed by esterification with POPAM **47** to afford a yield of 96%, the desired conjugate ENGA-L07E **57**.



Reagents : a- 1- NaBH₄, MeOH, 99%; 2- Ac₂O, DMAP, pyridine, 96%; b- NH₂OH.HCl, pyridine, 95%; c- SOCl₂, dioxane, 62%; d- LiOH, MeOH, 100%; e- POPAM **47**, DCC, DMAP, CH₂Cl₂, 96%.

Scheme 11. Synthesis of ENGA-L07E 57.

A similar way was used to synthesize the bislactam alkylator **61**, as depicted in Scheme 12. A Beckmann rearrangement of oximes **58** led to the desired bislactam **59** in a 75% yield.

It is interesting to note that all new synthesized steroidal lactams with POPAM **47** performed relatively very low acute toxicities, together with important antileukemic activity *in vitro* and *in vivo*, producing a high therapeutic ratio.

In 2017, Montiel-Smith et al.⁵⁹ reported the synthesis of steroidal lactams starting from the easily available cholesterol and pregnenolone. A new procedure for the synthesis of a 6α -aza-B-homo steroidal lactam analog of vespertilin, starting from diosgenin, was also described.

The synthesis of derivative **62** has been previously reported⁶⁰ from cholesterol by a synthetic route of five steps and polluting oxidants were used. Herein the formation of compound **62** was accomplished, in just one step, as depicted in Scheme 13 and the best yield of 65% was observed using 8 equivalents of BF₃·OEt₂ and NaNO₂ and a 3:1 (v/v) Ac₂O-AcOH ratio.



Reagents : a- NH₂OH.HCl, pyridine, 50%; b- SOCl₂, dioxane, 75%; c- LiOH, MeOH, 93%; d- POPAM **47**, DCC, DMAP, CH₂Cl₂, 70%.

Scheme 12. Synthesis of ENGA-DL02E 61.



 $Reagents: a-NaNO_2, BF_3.Et_2O, AcOH; b-Na_2CO_3, CH_2CI_2/MeOH, reflux; c-SOCI_2, dioxane.$

Scheme 13. Synthesis of steroidal lactam 64.

The new protocol described above for cholesterol was applied with pregnenolone **65** possessing a ketone at C-20, as reported in Scheme 14. Using the same conditions, the steroidal derivative **66** was isolated without alteration of the carbonyl group in a 77% yield. In the next step, the (*Z*)-oxime **67** was obtained by treatment of compound **66** with sodium carbonate Na₂CO₃. Finally, the Beckmann rearrangement of steroidal oxime **67** was successfully performed using SOCl₂ in dioxane and the desired B-homo steroidal lactam **68** was isolated in only 3 steps, with an overall yield of 27%.



Reagents : a- NaNO₂, BF₃.Et₂O, AcOH; b- Na₂CO₃, CH₂Cl₂/MeOH, reflux; c- SOCl₂, dioxane.

Scheme 14. Synthesis of steroidal lactam 68.

Then, the authors used the same conditions to afford steroidal lactam **64** from oxime **63**, as depicted in Scheme 15.

Firstly, the nitroimino derivative **70a** was synthesized in a poor yield (22% yield) by a nitrosation reaction of diosgenin acetate **69a** using the conditions described by Iglesias.⁶¹ Indeed, three side products were also obtained. The reaction was performed in AcOH with NaNO₂ (10 equiv) and BF₃·OEt₂ (10 equiv) and the temperature (15–20 °C) was controlled over 4 h. No purification of the crude products was done to prevent the degradation of the side chain promoted by the silica gel and alumina.⁶¹ A treatment of the mixture of compounds **70a** and **71a** was performed with NH₂OH·HCl and NaOAc in refluxing ethanol to convert the ketone group and the nitroimino moieties of the main compounds into oximes.⁶² The same treatment was applied to the mixture of derivatives **70b** and **71b**. The dioxime **72a** or **72b** was thus isolated without affecting the other functionalities (Scheme 14). In the next step, **72a** or **72b** was submitted under protic acidic conditions to a second-order Beckmann rearrangement to give in both cases vespertiline oxime **73**. The use of the mixture CF₃COOH, Ac₂O, BF₃·OEt₂ at room temperature led to the best results in a very short reaction time (20 min.) and to the acetylation of the hydroxyl group at C-3 in compound **72b**. Next, treatment of compound **73** with Na₂CO₃ in a 1:1 MeOH/CH₂Cl₂ refluxing solution led to the (*Z*)-oxime **74** without the ester group at C-5. In a final step,

the desired lactam **75** was isolated in a good yield of 70% using the same conditions described above for the synthesis of B-homo steroidal lactams **64** and **68**.

A panel of six human solid tumor cell lines was used to test the antiproliferative activity of these compounds. The authors observed that the most potent of all the derivatives assayed was compound **61a**, displaying slightly better GI50 values against WiDr and T-47D cancer cells when compared to cisplatin. A moderate activity in the cell lines screened was determined for the steroidal lactams **68** and **75**.



Reagents : a- NaNO₂, BF₃.Et₂O, AcOH; b- NH₂OH.HCl, EtOH, reflux; c- CF₃COOH, Ac₂O, BF₃.Et₂O; d- Na₂CO₃, CH₂Cl₂/MeOH, reflux; e- SOCl₂, dioxane.

Scheme 15. Synthesis of steroidal lactam 75.

In 2020, Liu et al.⁶³ reported the synthesis of biologically relevant steroidal spiro β -lactams from dienamides through the cascade 4-endo *N*-cyclization/aerobic oxidation sequence.

The synthetic pathway is depicted in Scheme 16. Dienamides **77** were prepared from α, α -dicyanoalkene **76**, and aldehydes through the cascade reactions under mild conditions.⁶⁴ Then, treatment of compound **66** in the presence of NaH in THF at 0 °C under air atmosphere led exclusively to the unexpected steroidal spiro β -lactams **78** within 1 h. Surprisingly, an additional hydroxyl group was also introduced at the α position of the β -lactam ring *via* the base-promoted aerobic oxidation. The above reactions proceeded selectively through the 4-endo *N*-cyclization in the presence of NaH, followed by the base-mediated aerobic oxidation under air. No 6-endo *N*-cyclization products, namely 2-piperidinones were observed. This approach has several advantages such as short reaction time and mild reaction conditions.



 $R = H, CI, Br, F, NO_2$

Reagents : a- ArCHO, cascade reaction;⁶⁴ b- NaH, THF, 0 °C, under air.

Scheme 16. Synthesis of steroidal β -lactam 78 from dienamide 77.

In 2021, Hernandez-Linares et al.⁶⁵ reported the synthesis of new steroidal lactams **82** and **87** from diosgenin **79** (Scheme 17 and 18).

In the case of the steroidal derivative **71**, the synthetic pathway is depicted in Scheme 17. Firstly, oxidation of diosgenin with PCC and calcium carbonate in dichloromethane led to the α , β -unsaturated compound **80** with a slightly improved yield (90%) compared to that described.⁶⁶ In the next step, this latter was oxidized using the oxidant reagent potassium permanganate-sodium periodate.⁶⁷ Thus, the use of permanganate provided the formation of 1,2-diol, which carbon-carbon bond was then cleaved to give a carboxylic acid group and a ketone, without degradation, leading to the desired derivative **81**.

In the final step, two techniques for the ring closure were used to obtain the targeted steroidal derivative **82**. In the first case, following the methodology reported by Jiang,⁶⁷ a conventional N source, ammonium sulfate in refluxing glacial acetic acid for 4 h, was attempted and the overall yield was improved from 39% to 63%. In the second case, focused microwave irradiation (FMWI)⁶⁷ was tried with ammonium acetate as a reagent, for 3 min, leading to a much higher yield of 75%, than that reported in previous works of 19%. Finally, the authors succeeded in increasing the yield to 94%, using 1 eq. of compound **70** with 3.8 eq. of ammonium acetate in acetic acid. The use of DMF instead of acetic acid decreased the yield to 71% and no reaction can be observed with ammonium hydroxide as an N source.



Reagents : a- PCC, CaCO₃, CH₂Cl₂, reflux, 5h; b- KMnO₄, NaIO₄, NaCO₃, *t*-BuOH/H₂O, 3.5 h, 89%; c- Method A: NH₄SO₄, AcOH, reflux, 4 h; Method B: NH₂OAc, AcOH, reflux, 4h; Method C: NH₂OAc, AcOH, MW, 180 °C, 3 min, 94%;

Scheme 17. Synthesis of steroidal lactam 82.

An oxidation reaction was tested with CrO₃ to obtain the lactam moiety in ring B, but the yield was low. The best yield of the allylic oxidation reaction was observed starting with diosgenin acetate as substrate and using the Collins reagent.⁶⁸ Thus, using compound **84**, which contains a ketone group, as a precursor, a ring expansion was done leading to the intermediary oxime **86**. In the next step, a Beckmann rearrangement was performed with SOCl₂ in THF, to provide the targeted 7-azasteroid **87** as depicted in Scheme 18.

Moreover, the by-product **85** with a carboxylic group, was isolated in addition to **84**. The treatment of enone **84** with NH₂OH.HCl under reflux in an ethanol and sodium acetate solution for two hours led to the corresponding oxime with a yield of 91%. One equivalent of the enone for 1.5 eq. of NH₂OH.HCl was used to obtain the best yield. Finally, the reaction of derivative **86** with a solution of thionyl chloride in THF for 4 h at 0 $^{\circ}C^{69}$ provided the steroidal lactam-type enamide **87**.

A specificity to different types of cellular receptors in cancer cell lines tested, as well as significant antiproliferative activities, were observed for some of the synthesized compounds, depending on the structure and conformation. The steroidal lactam **82** inhibits cancer cells lines with the PR and ER α without showing cytotoxicity for lymphocytes of PHB; however, the derivative seco-ketoacid **81** does not inhibit cells with the receptors ER α , ER β , and PR.

Moreover, it was observed that the steroidal lactam **87** inhibits the proliferation in MDA-MB-231 cell lines that do not express ER α , PR, or HER2 receptors, with IC50 values lower than 0.093 µg/mL, showing that the route through which that bind to cells is different.



Reagents : a- Ac₂O, DMAP, reflux, 1 h, 85%; b- Method A: CrO₃, AcOH/H₂O, 0 °C, 3 h; Method B: Collins reagent, CH₂Cl₂, r.t., 12 h, 71%; c- NH₂OH.HCl, AcONa, EtOH, reflux, 2 h, 91%; d- SOCl₂, THF, 0 °C, 4 h, 52%;

Scheme 18. Synthesis of steroidal lactam 87.

3. Conclusions

The lactam moiety and the steroid nucleus are prevalent in drug molecules and natural products, the derivatives possessing such fragments always present diverse and interesting biological profiles. Presented here is up-to-date literature on the syntheses of steroid lactams reported during the last years. Several of these syntheses may be useful, and in most cases reporting the cytotoxicity of the tested compounds, there seems to be a link with the presence of the lactam moiety in the steroid skeleton. Currently, interest in steroids and related molecules continues because of the emerging bioactivity and structural diversity inherent in this class of compounds.

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5. References

- 1. Zeelen, J. F. *Medicinal chemistry of steroids*; Elsevier: Amsterdam, Netherlands, 1990.
- 2. Trager, L. F. Steroidhormone; Springer: Berlin, 1977.
- 3. Biellmann, J. F. *Chem. Rev.* **2003**, *103*, 2019. http://doi.org/10.1021/cr020071b
- 4. Nising, C. F.; Bräse, S. *Angew. Chem. Int. Ed.* **2008**, *47*, 9389. https://doi.org/10.1002/anie.200803720
- 5. Hanson, J. R. *Nat. Prod. Reports* **2010**, *27*, 887. https://doi.org/10.1039/C001262A
- Wilson, R. M.; Danishefsky, S. J. J. Org. Chem. 2006, 71, 8329. <u>https://doi.org/10.1021/jo0610053</u>
- 7. Kumar, K.; Waldmann, H. *Angew. Chem. Int. Ed.* **2009**, *48*, 3224. https://doi.org/10.1002/anie.200803437
- Maurin, P.; Ibrahim-Ouali, M., Santelli, M. Eur. J. Org. Chem. 2002, 1, 151. https://doi.org/10.1002/1099-0690(20021)2002:1<151::AID-EJOC151>3.0.CO;2-F
- 9. Maurin, P.; Ibrahim-Ouali, M., Santelli, M. *Tetrahedron Lett.* **2001**, *42*, 847. https://doi.org/10.1016/S0040-4039(00)02119-5
- 10. Oumzil, K.; Ibrahim-Ouali, M.; Santelli, M. *Synlett* **2005**, *11*, 1695. <u>https://doi.org/10.1055/s-2005-869879</u>
- 11. Cachoux, F.; Ibrahim-Ouali, M.; Santelli, M. *Tetrahedron Lett.* **2001**, *42*, 843. https://doi.org/10.1016/S0040-4039(00)02118-3
- 12. Oumzil, K.; Ibrahim-Ouali, M.; Santelli, M. *Tetrahedron Lett.* **2005**, *46*, 5783. <u>https://doi.org/10.1016/j.steroids.2006.06.003</u>
- 13. Cachoux, F.; Ibrahim-Ouali, M.; Santelli, M. *Synlett* **2000**, *3*, 418. <u>https://doi.org/10.1055/s-2000-6534</u>
- 14. Oumzil, K.; Ibrahim-Ouali, M.; Santelli, M. *Tetrahedron* **2005**, *61*, 9405. https://doi.org/10.1016/j.tet.2005.07.026

- 15. Ibrahim-Ouali, M. *Tetrahedron Lett.* **2009**, *50*, 1607. <u>https://doi.org/10.1016/j.tetlet.2009.01.107</u>
- 16. Ibrahim-Ouali, M. *Tetrahedron Lett.* **2010**, *51*, 3610. https://doi.org/10.1016/j.tetlet.2010.05.008
- 17. Ibrahim-Ouali, M.; Romero, E. *Steroids* **2012**, *77*, 157. <u>https://doi.org/10.1016/j.steroids.2011.11.003</u>
- 18. Cachoux, F.; Ibrahim-Ouali, M.; Santelli, M. *Synth Commun* **2002**, *32*, 3549. <u>https://doi.org/10.1081/SCC-120014965</u>
- 19. Oumzil, K.; Ibrahim-Ouali, M.; Santelli, M. *Steroids* **2006**, *71*, 886. https://doi.org/10.1016/j.steroids.2006.06.003
- 20. Oumzil, K.; Ibrahim-Ouali, M.; Santelli, M. *Tetrahedron Lett.* **2005**, *46*, 5799. https://doi.org/10.1016/j.tetlet.2005.06.150
- 21. Ibrahim-Ouali, M.; Romero, E.; Bouleghlem, H. *Tetrahedron* **2011**, *67*, 3668. <u>https://doi.org/10.1016/j.tet.2011.03.080</u>
- 22. Ibrahim-Ouali, M.; Zoubir, J.; Romero, E. *Tetrahedron Lett.* **2011**, *52*, 7128. <u>https://doi.org/10.1016/j.tetlet.2011.10.108</u>
- 23. Ibrahim-Ouali, M.; Bouleghlem, H.; Aouf, N. E. *Tetrahedron Lett.* **2012**, *53*, 1859. <u>https://doi.org/10.1016/j.tetlet.2012.02.005</u>
- 24. Ibrahim-Ouali, M.; Romero, E.; Hamze, K. *Steroids* **2012**, *77*, 1092. https://doi.org/10.1016/j.steroids.2012.04.004
- 25. Ibrahim-Ouali, M.; Romero, E. Steroids 2013, 78, 651. https://doi.org/10.1016/j.steroids.2013.03.004
- 26. Ibrahim-Ouali, M.; Hamze, K. *Steroids* **2014**, *80*, 102. https://doi.org/10.1016/j.steroids.2013.12.002
- 27. Ibrahim-Ouali, M. *Steroids* **2015**, *98*, 9. https://doi.org/10.1016/j.steroids.2015.02.014
- 28. Ibrahim-Ouali, M. *Steroids* **2006**, *71*, 1025. https://doi.org/10.1016/j.steroids.2006.09.006
- 29. Ibrahim-Ouali, M. *Steroids* **2007**, *72*, 475. https://doi.org/10.1016/j.steroids.2007.03.004
- 30. Ibrahim-Ouali, M.; Rocheblave, L. *Steroids* **2008**, *73*, 375. https://doi.org/10.1016/j.steroids.2007.12.013
- 31. Ibrahim-Ouali, M. *Steroids* **2008**, *73*, 775. https://doi.org/10.1016/j.steroids.2008.04.005
- 32. Ibrahim-Ouali, M. *Steroids* **2009**, *74*, 133. https://doi.org/10.1016/j.steroids.2008.10.012
- 33. Ibrahim-Ouali, M.; Dumur, F. *Arkivoc* **2017**, 202. https://doi.org/10.24820/ark.5550190.p009.986
- 34. Ibrahim-Ouali, M.; Dumur, F. *Arkivoc* **2019**, 304. https://doi.org/10.24820/ark.5550190.p010.988
- 35. Ibrahim-Ouali, M.; Dumur, F. *Arkivoc* **2021**, 130. https://doi.org/10.24820/ark.5550190.p011.543
- 36. Ibrahim-Ouali, M.; Dumur, F. *Arkivoc* **2021**, 300. https://doi.org/10.24820/ark.5550190.p011.614

- 37. Ibrahim-Ouali, M.; Dumur, F. Arkivoc **2021**, 471. https://doi.org/10.24820/ark.5550190.p011.512
- 38. Ibrahim-Ouali, M.; Dumur, F. *Arkivoc* **2022**, 140. https://doi.org/10.24820/ark.5550190.p011.734
- 39. Catsoulacos, P.; Politis, D.; Wampler, G.L. *Cancer Chemother. Pharmacol.* **1983**, *10*, 129. https://doi.org/10.1007/BF00446225
- 40. Metcalf, B.W.; Levy, M.A.; Holt, D.A. *Trends Pharm. Sci.* **1989**, *10*, 491. https://doi.org/10.1016/0165-6147(89)90048-5
- 41. Liang, T.; Rasmusson, G.H.; Brooks, J.R. *J. Steroid Biochem.* **1983**, *19*, 385. https://doi.org/10.1016/s0022-4731(83)80051-x
- Rasmusson, G.H.; Reynolds, G.F.; Steinberg, N.G.; Walton, E.; Pate, G.F.; Liang, T.; Cascieri, M.A.; Cheung, A.H.; Brooks, J.R.; Berman, C. J. Med. Chem. 1986, 9, 2298. https://doi.org/10.1021/jm00161a028
- 43. Georgianna, S.H.; Kozarich, J.W. *Curr. Opin. Chem. Biol.* **1997**, *1*, 254. <u>https://doi.org/10.1016/S1367-5931(97)80017-8</u>
- 44. Singh, R.; Panda, G. *Tetrahedron* **2013**, *69*, 2853. https://doi.org/10.1016/j.tet.2013.02.018
- 45. Huang, Y.; Cui, J.; Chen, S.; Gan, C.; Zhou, A. *Steroids* **2011**, *76*, 1346. <u>https://doi.org/10.1016/j.steroids.2011.06.013</u>
- 46. Cui, J.G.; Fan, L.; Huang, Y.M.; Xin, Y.; Zhou, A.M. *Steroids* **2009**, *74*, 989. https://doi.org/10.1016/j.steroids.2009.07.009
- 47. Huang, Y.; Cui, J.; Zheng, Q.; Zeng, C.; Chen, Q.; Zhou, A. *Steroids* **2012**, *77*, 829. http://dx.doi.org/10.1016/j.steroids.2012.04.016
- 48. 48.Huang, Y.; Cui, J.; Chen, S.; Lin, Q.; Song, H.; Gan, C.; Su, B.; Zhou, A. *Mar. drugs* **2014**, *12*, 1715. <u>https://doi.org/10.3390/md12041715</u>
- 49. Cui, J.G.; Fan, L.; Huang, L.L.; Liu, H.L.; Zhou, A.M. *Steroids* **2009**, *74*, 62. https://doi.org/10.1016/j.steroids.2008.09.003
- 50. Cui, J.G.; Fan, L.; Huang, Y.M.; Xin, Y.; Zhou, A.M. *Steroids* **2009**, *74*, 989. <u>https://doi.org/10.1016/j.steroids.2009.07.009</u>
- 51. Cui, J.; Lin, Q.; Huang, Y.; Gan, C.; Yao, Q.; Wie, Y.; Xiao, Q.; Kong, E. *Med. Chem. Res.* **2015**, *24*, 2906. https://doi.org/10.1007/s00044-015-1347-3
- 52. Trafalis, D.; Geromichalou, E.; Dalezis, P.; Nikoleousakos , N.; Sarli , V. Steroids **2016**, *115*, 1. http://dx.doi.org/10.1016/j.steroids.2016.07.009
- 53. Camoutsis, C.; Catsoulacos, P. *J. Heterocyclic Chem.* **1983**, *20*, 1093. <u>https://doi.org/10.1021/ja01126a070</u>
- 54. Suginome, H.; Kaji, M.; Yamada, S. *J. Chem. Soc., Perkin Trans.* 1 **1988**, 321. https://doi.org/10.1039/P19880000321
- 55. Yao, Z.; Xu, Y.; Zhang, M.; Jiang, S.; Nicklaus, M.C.; Liao, C. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 475. https://doi.org/10.1016/j.bmcl.2010.10.112
- 56. Zhao, Q.; Li, Z. *Steroids* **1994**, *59*, 190. <u>https://doi.org/10.1016/0039-128X(94)90027-2</u>
- 57. Morreal, C.E. *Steroids* **1966**, *8*, 671. https://doi.org/10.1016/0039-128X(66)90007-9
- 58. Norymberski, J.K.; Woods, G.F. J. Chem. Soc. 1955, 3426.

https://doi.org/10.1039/JR9550003426

- Martínez-Pascual, R.; Meza-Reyes, S.; Vega-Baez, J. L.; Merino-Montiel, P.; Padrón, J. M.; Mendoza, Á.; Montiel-Smith, S. Steroids 2017, 122, 24. <u>https://doi.org/10.1016/j.steroids.2017.03.008</u>
- 60. Duddeck, H.; Frelek, J.; Szczepek, W.J.; Rodewald, W.J. Bull. Chem. Soc. Ethiop. 1987, 1, 18.
- Y. López, K.M. Ruíz-Pérez, R. Yépez, R. Santillan, M. Flores-Alamo, M.A. Iglesias-Arteaga. Steroids 2008, 73, 657.

http://dx.doi.org/10.1016/j.steroids.2008.02.003

- 62. Ranise, A.; Bondavalli, F.; Schenone, P.; Mugnoli, A.; Pani, M. J. Chem. Soc., Perkin Trans. 1 **1990**, 3053. http://dx.doi.org/10.1039/P19900003053.
- 63. Fanga, Y.; Liaoa, G.; Guoc, H.; Yub, B.; Hong-Min, Liu. *Steroids* **2020**, *159*, 108635. https://doi.org/10.1016/j.steroids.2020.108635
- 64. Yu, B.; Shi, X.J.; Ren, J.L.; Sun, X.N.; Fang, Y.; Ye, X.W.; Wang, M.M.; Yu, Q.; Liu, H.M. *Eur. J. Med. Chem.*2013, 66, 171. https://doi.org/10.1016/j.ejmech.2013.05.035
- Martínez-Gallegos, A.A.; Guerrero-Luna, G.; Ortiz-Gonzalez, A.; Cardenas-García, M.; Bernes, S.; Hernandez-Linares, M. G. Steroids 2021, 166, 108777. https://doi.org/10.1016/j.steroids.2020.108777
- 66. Shawakfeh, K.Q.; Al-Said, N.H. *Steroids* **2011**, *76*, 232. https://doi.org/10.1016/j.steroids.2010.10.002.
- 67. Jiang, Z.X.; Ye, J.Q.; Jiang, L.; Zhao, Y.S. *Steroids* **2005**, *70*, 690. https://doi.org/10.1016/j.steroids.2005.03.009.
- Hamid, A.A.; Kaushal, T.; Ashraf, R.; Singh, A.; Chand Gupta, A.; Prakash, O.; Sarkar, J.; Chanda, D.; Bawankule, D.U.; Khan, F.; Shanker, K.; Aiyelaagbe, O.O.; Negi. A.S. Steroids 2017, 119, 43. <u>https://doi.org/10.1016/j.steroids.2017.01.001.</u>
- 69. Zhi, X.; Zhang, Y.; Huang, J.; Xu, H. *Sci. Rep.* **2017**, *7*. https://doi.org/ 10.1038/s41598-017-04136-3.

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