2-Aminopteridin-4-one derivatives from 2, 3-dicyanopyrazine

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Abstract

2-Allylamino-3-cyanopyrazine (3) was transformed into 1-allylpteridin-4-one-2-thiones (4) by the reaction with isothiocyanates, and the thiones 4 were thermally converted into 2-allylthiopteridin-4-ones (5) by 3,3-sigmatropic rearrangement. The allylthio group of 5 was replaced by a butylamino group to give 2-butylaminopteridin-4-ones (6).

Keywords: Pteridine synthesis, nucleophilic substitution of pyrazine, S-Cope rearrangement

Introduction

2-Aminopteridin-4-one (1) constitutes a core structure in the commonly occurring coenzymes such as biopterin, folic acid, and methanopterin. Previously, we have investigated reactions of 2,3-dicyanopyrazine, which was conveniently prepared from diaminomaleonitrile and glyoxal, and its derivatization to pteridine derivatives. 2,3-Dicyanopyrazine (2) is easily converted into 2-allylamino-3-cyanopyrazine (3) by the simple treatment with allylamine. In this paper we would like to report the synthesis of 2-aminopteridin-4-one derivatives, starting from 2,3-dicyanopyrazine (2).

Results and Discussion

2-Allylamino-3-cyanopyrazine (3) was conveniently prepared by the treatment of 2,3-dicyanopyrazine (2) with allylamine⁶ and then the cyanopyrazine 3 was converted to 1-allyl-3-phenyl- (4a), 1-allyl-3-butyl- (4b), or 1-allyl-3-methyllpteridin-4-one-2-thione (4c) in

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69-88% yields by the reactions with the corresponding isothiocyanates followed by hydrolysis of the intermediate imines.

Pteridinethione derivatives **4a**, **4b** and **4c** were heated to 180-198°C to cause the 3,3-sigmatropic rearrangement, and the thermal equilibration produced allylthio derivatives **5a**, **5b** and **5c** as components of the equilibrium mixtures (40%, 23%, and 21% respectively) along with the intact starting materials. Thermal treatment of the isolated products **5a**, **5b** and **5c** gave the same equilibrium mixtures of **4**'s and **5**'s by the re-equilibration, and therefore this transformation is reversible. Unfortunately, the equilibria between N-allyl derivatives **4**'s and S-allyl derivatives **5**'s favor the former, but the starting materials **4**'s can be efficiently recovered and subjected to another thermal isomerization. The treatment of compounds **5** with butylamine substitutes the allylthio group with butylamine to produce 2-butylamino-3-phenyl- (**6a**), 2-butylamino-3-butyl-(**6b**), and 2-butylamino-3-methylpteridin-4-ones (**6c**) in 60-85% yields. Other alkylamines react in the same manner though only butylamine is shown here as an example (Scheme 1).

4a - 6a (R=Ph); 4b - 6b (R=Bu); 4c - 6c (R=Me)

Scheme 1. Derivation of aminopteridin-4-ones by S-Cope rearrangement.

Structures of thiones **4a-4c** and the allylthio derivatives **5a-5c** were assigned straightforwardly by the NMR signals due to the protons on the pyrazine ring and by the appropriate shift of the signals of the allylic protons due to the rearrangement, N-CH₂ 5.58-5.59 for **4**'s and S-CH₂ 3.96-4.08 for **5**'s. The substitution **5** \rightarrow **6** is evident from the replacement of the signals due to the allylthio group by those due to the butylamino group.

The removal of the allyl group from thiones **4** was attempted by the previously reported methods such as palladium catalysis, hydrogen abstraction by thiyl radical, and allyl group transfer to methyl chloroformate. All these attempts to remove the allyl group at this stage were

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unsuccessful probably due to the steric hindrance for the approach of a reagent. The present thio-Cope rearrangement is a promising method for the removal of N-allyl group and the introduction of amino group by the nucleophilic substitution is efficient. The procedure reported in this paper is a convenient and simple route to the synthesis of 2-aminopteridin-4-one derivatives, although the appropriate structural modifications are required for the favorable S-Cope rearrangement of thiones 4.

Experimental Section

Synthesis of 1-allylpteridin-4-one-2-thiones (4a, 4b and 4c). In a nitrogen-flushed flask were placed sodium hydride (60% dispersion with mineral oil, 176 mg, 4.4 mmol) and 640 mg (4.0 mmol) of 2-allylamino-3-cyanopyrazine (3)⁶ dissolved in 20 ml of THF, and the mixture was stirred for 10 min at room temperature. Phenylisothiocyanate (810 mg, 6.0 mmol) was added dropwise to the mixture and the reaction mixture was stirred for 2.5 h at room temperature. After the addition of 2M HCl, washing with chloroform, and neutralization with aq. NaHCO₃, the product was extracted three times with chloroform. Conventional work up of the extract gave crude product of 1-allyl-3-phenylpteridin-4-imino-2-one. This imino derivative was hydrolyzed to 4a by refluxing for 1.0 h in 2 M HCl. The reaction mixture was extracted three times with chloroform after neutralization with saturated aq. NaHCO₃. The crude product was recrystallized from ethanol to give 1.04 g of 1-allyl-3-phenylpteridin-4-one-2-thione (4a) (88%). The thiones 4b and 4c were prepared by the same procedure in 77 and 69 % yields.

4a. Mp 197.5-198.8°. 1 H-NMR (400MHz, CDCl₃) δ 5.31(1H, dtd, J=10.2, 1.5 and 1.2Hz), 5.38(1H, dtd, J=17.5, 1.2 and 1.2Hz), 5.59(2H, ddd, J=5.6, 1.2 and 1.2Hz), 6.03(1H, ddt, J=17.5, 10.0 and 5.6Hz), 7.24(2H, m), 7.50(1H, m), 7.56(2H, m), 8.63(1H, d, J=2.2Hz), 8.76(1H, d, J=2.2Hz); 13 C-NMR (150MHz, CDCl₃) 50.8, 119.4, 128.1, 128.9, 129.5, 129.7, 130.0, 130.3, 141.5, 148.1, 148.4, 157.9, 178.0; IR (CHCl₃) 3012, 3007, 1710, 1579, 1547, 1475, 1428, 1385 cm⁻¹; MS(FAB) m/z=297 (M⁺ + H). Anal. C, 60.44; H, 3.75; N, 18.82. Calcd for $C_{15}H_{12}N_4OS$: C, 60.79; H, 4.08; N, 18.91.

4b. Mp 144.0-145.1°. 1 H-NMR (400MHz, CDCl₃) δ 0.99(3H, t, J=7.3Hz), 1.45(2H, tq, J=7.6 and 7.3Hz), 1.79(2H, tt, J=7.8 and 7.6Hz), 4.65(2H, t, J=7.8Hz), 5.27(1H, d, J=10.5Hz), 5.29(1H, d, J=17.1Hz), 5.58(2H, d, J=5.4Hz), 6.01(1H, ddd, J=17.1, 10.5 and 5.4Hz), 8.65(1H, d, J=2.2Hz), 8.70(1H, d, J=2.2Hz); 13 C-NMR (150MHz, CDCl₃) 13.7, 20.1, 28.2, 48.7, 50.5, 118.9, 129.2, 130.5, 141.5, 147.8, 147.9, 157.7, 176.8; IR (CHCl₃) 3014, 1700, 1549, 1482, 1438, 1420 cm⁻¹; MS(FAB) m/z=277(M⁺+H). Anal. C, 56.47; H, 5.66; N, 20.20. Calcd for C₁₃H₁₆N₄OS: C, 56.50; H, 5.84; N, 20.27.

4c. Mp 172.0-174.2°. ¹H-NMR (400MHz, CDCl₃) δ 3.94(3H, s), 5.27(1H, dtd, J=9.9, 1.5 and 1.0Hz), 5.30(1H, dtd, J=16.5, 1.5 and 1.0Hz), 5.59(2H, ddd, J=5.6, 1.5, and 1.5Hz), 6.01(1H, ddt, J=16.5, 9.9 and 5.6Hz), 8.67(1H, d, J=2.1), 8,73(1H, d, J=2.1Hz); ¹³C-NMR (150MHz, CDCl₃) 36,3, 50.6, 119.1, 129.0, 130.4, 141.6, 147.8, 148.0, 158.2, 177.2; IR (CHCl₃) 3015, 1705, 1550,

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1484, 1438, 1419 cm⁻¹; HRMS(FAB) m/z=235.0670. Calcd for $(C_{10}H_{10}N_4OS + H)$ m/z=235.0653. Anal. C, 51.21; H, 4.01; N, 23.62. Calcd for $C_{10}H_{10}N_4OS$: C, 51.27; H, 4.30; N, 23.91.

Thermal isomerization of 1-allylpteridin-4-one-2-thiones (4a, 4b and 4c) to 3-allylthio-pteridin-4-ones (5a, 5b and 5c). Crystalline thione **4a** (69 mg, 0.2 mmol) was placed in a glass tube and heated to the melting point (198°) for 2 h under argon atmosphere. Chromatography of the residue on silica gel eluted with hexane-ethyl acetate (2:1) yielded allylthio derivative **5a** in 40 % yield and mostly recovered **4a**. Thione **4b** was heated at 180° in refluxing o-dichlorobenzene for 48 h, and thione **4c** was heated at 190° for 20 h in o-dichlorobenzene using a sealed glass tube. Allylthio derivatives **5b** and **5c** were obtained from **4b** and **4c** in the same work up in 23% and 21% respectively.

5a. ¹H-NMR (400MHz, CDCl₃) δ 3.96(2H, ddd, J=5.1, 1.2 and 1.2Hz), 5.18(1H, dtd, J=10.0, 1.2 and 1.5Hz), 5.34(1H, dtd, J=17.0, 1.2 and 1.5Hz), 5.93(1H, ddt, J=17.0, 10.0 and 5.1Hz), 7.34(2H, m), 7.58(3H, m), 8.76(1H, d, J=2.2), 8.90(1H, d, J=2.2); ¹³C-NMR (150MHz, CDCl₃) 36.5, 119.9, 128.6, 129.2, 130.5, 130.8, 131.4, 131.9, 141.9, 143.9, 150.5, 160.8, 164.0: IR (CHCl₃) 3015, 1714, 1559, 1539, 1526, 1404 cm⁻¹; HRMS(FAB) m/z=297.0841. Calcd for (C₁₅H₁₂N₄OS + H) m/z=297.0812.

5b. 1 H-NMR (400MHz, CDCl₃) δ 1.00(3H, t, J=7.3), 1.47(2H, tq, J=7.6 and 7.3), 1.80(2H, tt, J=8.1 and 7.6), 4.08(2H, ddd, J=6.8, 1.2 and 1.2Hz), 4.19(2H, t, J=8.1Hz), 5.26(1H, dtd, J=10.0, 1.0 and 1.0Hz), 5.44(1H, dtd, J=17.1, 1.2 and 1.0Hz), 6.03(1H, ddt, 17.1, 10.0 and 6.8Hz), 8.72(1H, d, J=2.0Hz), 8.85(1H, d, J=2.0Hz); 13 C-NMR (150MHz, CDCl₃) 13.8, 20.3, 29.8, 35.6, 45.4, 120.0, 131.5, 131.6, 143.6, 150.1, 153.2, 160.5, 162.3; IR (CHCl₃) 3019, 3015, 1700, 1550, 1532, 1411 cm⁻¹; HRMS(FAB) m/z=277.1118. Calcd for (C₁₃H₁₆N₄OS + H) m/z=277.1123.

5c. ¹H-NMR (400MHz, CDCl₃) δ 3.69(3H, s), 4.09(2H, d, J=6.8), 5.26(1H, dd, J=9.8 and 1.5Hz), 5.45(1H, dd, J=16.6 and 1.5Hz), 6.03(1H, ddt, J=16.6, 9.8 and 6.8Hz), 8.73(1H, d, J=2.4Hz), 8.86(1H, d, J=2.4Hz); ¹³C-NMR (150MHz, CDCl₃) 31.5, 35.9, 120.3, 131.6, 131.8, 143.9, 144.0, 150.5, 161.1, 163.1; IR (CHCl₃) 3006, 1700, 1550, 1534, 1419 cm⁻¹; HRMS(FAB) m/z=235.0672. Calcd for (C₁₀H₁₀N₄OS + H) m/z=235.0653.

Synthesis of 2-butylaminopteridin-4-ones (6a, 6b and 6c). 2-Allylthio-3-phenylpyteridin-4-one (5a) (20 mg, 0.068 mmol) was placed in a nitrogen-flushed flask and treated with 0.14 ml (1.4 mmol) of butylamine dissolved in 0.5 ml of THF and the mixture was refluxed for 24 h. Concentration and chromatography of the reaction mixture on silica gel eluted with ethyl acetate-ethanol (10:1) gave 2-butylamino-3-phenylpteridin-4-one (6a) in 60% yield. The same procedure gave 6b and 6c from 5b and 5c in 80 and 85% yield, respectively.

6a. ¹H-NMR (400MHz, CDCl₃) δ 0.91(3H, t, J=7.3Hz), 1.38(2H, q, J=7.3Hz), 1.59(2H, tt, J=7.1 and 7.3Hz), 3.47(2H, td, J=7.1 and 5.4Hz), 4.35(1H, bs, NH), 7.25-7.29(2H, m), 7.52-7.61(3H, m), 8.43(1H, d, J=2.2Hz), 8.65(1H, d, J=2.2Hz); ¹³C-NMR (150MHz, CDCl₃) 13.7, 20.0, 31,1, 42.0, 128.6, 129.9, 139.6, 131.0, 133.7, 140.3, 150.0, 151.9, 156.0, 161.1; IR (CHCl₃) 3434, 3000, 1701, 1564, 1412 cm⁻¹; HRMS(FAB) m/z=296.1519. Calcd for (C₁₆H₁₇N₅O + H) m/z=296.1511.

6b. ¹H-NMR (400MHz, CDCl₃) δ 0.98(3H, t, J=7.3Hz), 0.99(3H, t, J=7.3Hz), 1.44(2H, tq, J=7.1 and 7.3), 1.46(2H, tq, J=7.1 and 7.3Hz), 1.70(2H, tt, J=7.3 and 7.1Hz), 1.73(2H, tt, J=7.6 and

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7.1Hz), 3.68(2H, td, J=7.3 and 5.4Hz), 4.10(2H, t, J=7.6Hz), 5.10(1H, bs, NH), 8.47(1H, d, J=2.0Hz), 8.69(1H, d, J=2.0Hz); 13 C-NMR (150MHz, CDCl₃) 13.7, 13.8, 20.1, 20.2, 29.4, 31.3, 41.5, 42.3, 129.1, 140.1, 149.9, 151.7, 155.4, 161.4; IR (CHCl₃) 3480, 3014, 1700, 1580, 1540, 1471 cm⁻¹; HRMS(FAB) m/z=276.1801. Calcd for (C₁₄H₂₁N₅O + H) m/z=276.1824.

6c. ¹H-NMR (400MHz, CDCl₃) δ 0.99(3H, t, J=7.3Hz), 1.46(2H, tq, J=7.4 and 7.3Hz), 1.71(2H, tt, J=7.3 and 7.3Hz), 3.58(3H, s), 3.69(2H, td, J=7.3 and 5.3Hz), 3.86(1H, bs, NH), 8.49(1H, d, J=2.2Hz), 8.71(1H, d, J=2.2Hz); ¹³C-NMR (150MHz, CDCl₃) 13.7, 20.1, 28.1, 31.2, 42.4, 128.9, 140.4, 150.0, 152.1, 155.3, 161.4; IR (CHCl₃) 3474, 3013, 3001, 1696, 1583, 1565, 1476 cm⁻¹; HRMS(FAB) m/z=234.1372. Calcd for (C₁₁H₁₅N₅O + H) m/z=234.1355.

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