

Synthesis, azo-hydrazone tautomerism and antitumor screening of N-(3-ethoxycarbonyl-4,5,6,7-tetrahydro-benzo[b]thien-2-yl)-2-arylhydrazono-3-oxobutanamide derivatives

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Abstract

A series of new *N*-(3-ethoxycarbonyl-4,5,6,7-tetrahydrobenzo[b]thien-2-yl)-2-aryl hydrazono-3-oxobutanamide derivatives **4a-i** were synthesized and the acid dissociation constants (pK_{as}) for the series prepared were determined and correlated by the Hammett-type equation using the enhanced substituent constants σ_x^- . The results of such correlation together with the spectral data indicated that the studied compounds exist predominantly in the hydrazone tautomeric form. Some of these derivatives exhibit high antitumor activity.

Keywords: Synthesis, coupling, azo-hydrazone tautomerism, benzo[b]thiophene

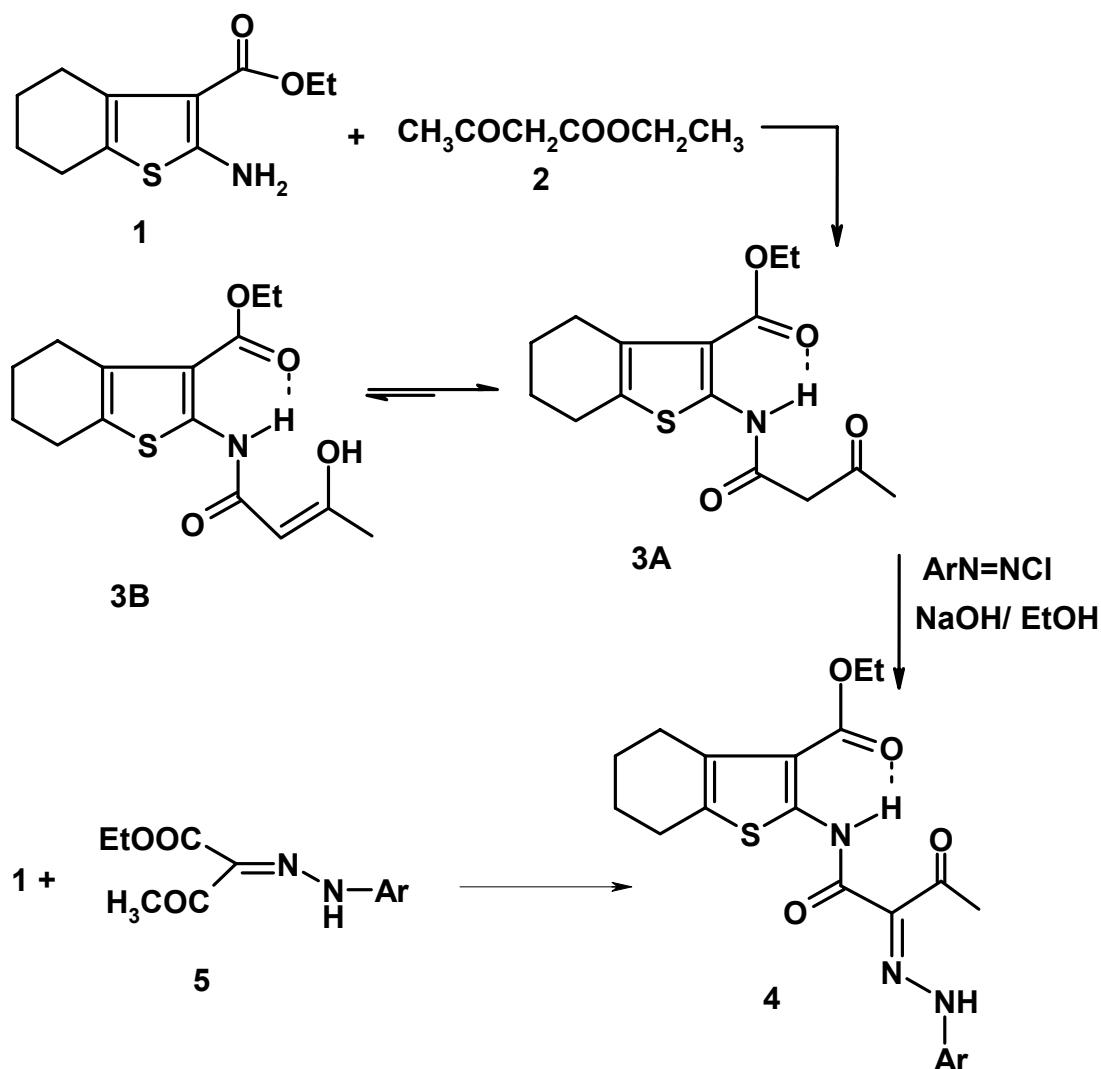
Introduction

Azo dyes are versatile compounds that have wide applications in industry and photodynamic therapy as well as photosensitive species in photographic or electrophotographic systems and are dominant organic photoconductive materials.¹ Furthermore, thiophenes and their fused derivatives have shown diverse pharmacological activities including antibacterial,^{2,3} antifungal,³ immunomodulatory,⁴ antidiabetic,⁵ antiinflammatory,⁶ antiviral activities^{7,8} and anticancer agent.⁹ Therefore, it was thought to be of interest to combine two of the above-mentioned azo dye and thiophene ring together in a molecular framework to give dye with high pharmaceutical potentiality.

Results and Discussion

The required starting active methylene compound **3** was previously prepared¹⁰ as a mixture in 15-20% yield, but in this study we synthesized it by the condensation of compound **1** with ethyl acetoacetate **2** at 160 °C for 30 min. to give compound **3** in 95% yield (Scheme 1). Compound **3**

can exist in one of two tautomeric structures **3A** or **3B**. The IR spectral data seem to be more consistent with the ketone structure **3A**, since the IR spectra revealed three bands at 1709, 1682, 1666 cm⁻¹ corresponding to the stretching vibration of the ester, acetyl and amide carbonyl groups, respectively. The low stretching vibrations of ester carbonyl group was due to the hydrogen bonding with the amide NH. In addition, the ¹H NMR spectra show the presence of NH singlet signal at 11.43 ppm and the absence of singlet signal at 5.70 ppm for =CH. These findings indicate that compound **3** exist predominantly in the ketone form **3A** (Scheme 1).



Scheme 1. Synthesis of compound **4a-i**.

Reaction of **3** with diazotized aniline derivatives in aqueous ethanol in the presence of sodium hydroxide afforded the respective arylazo derivatives **4a-i** (Scheme 1). The mass spectra of the latter products revealed the molecular ion peaks at the expected m/z values with relative intensities varying from 40 to 100% and their elemental analysis data are consistent with their assigned structure. As shown in Figure 1, compounds **4** can have one or more of three possible tautomeric structures **4A-C** namely, keto-hydrazone **4A**, hydroxy-azo **4B** and CH-azo **4C**. Of these forms, structure **4A** seems to be the form of choice as it is consistent with their electronic absorption and ¹H NMR spectra. For example, like typical hydrazones,^{11,12} the electronic absorption spectra of **4** in dioxane revealed in each case two characteristic absorption bands in the regions 394-411 and 243-281 nm (Table 1). Furthermore, the spectrum of **4d**, taken as a typical example of the series studied was recorded in solvents of different polarities. The spectra obtained showed little, if any, shift (Table 1). The small shifts in λ_{max} of **4d** in different solvents are due to solute-solvent interaction. In agreement with this conclusion is the observation that the spectra of arylhydrazones derived from the reaction of quinones with *N*-alkyl-*N*-phenylhydrazine, unlike those of *o*- and *p*-hydroxyazo compounds, are largely independent of the solvent polarity.¹² This finding, while it excludes the azo tautomeric forms **4B** and **4C**, indicates that each of compounds **4** exists in one tautomeric form, namely **4A** (Figure 1). This conclusion was confirmed by the ¹H NMR and IR spectra of the studied compounds **4**. Their ¹H NMR spectra revealed no signals at 5.3 ppm for CH of tautomeric form **4C** and their IR spectra revealed three carbonyl absorption bands in regions 1701-1671, 1666-1651 and 1655-1624 cm⁻¹ characteristic for ester, acetyl and amide carbonyl groups, respectively. The observed low frequency of all of these carbonyl groups may be the result of the intramolecular H-bonding.

The assigned structure **4** was confirmed by an alternative synthesis of the products **4a** and **4d** as typical examples of the series prepared. Thus, treatment of **1** with each of the compounds **5a** and **5d** in dioxan in presence of triethylamine gave the products **4a** and **4d** that proved identical in all respects (m.p., mixed m.p., IR, UV) with those obtained above from the coupling of **3** with the respective diazotized anilines (Scheme 1).

To provide further evidence for the tautomeric form **4A** assigned to the studied hydrazone products **4**, their acid dissociation constants, pK_{as} , of the series prepared were determined and their correlation by the Hammett equation was examined.¹³ The acid dissociation constants for the series **4a-i** were determined potentiometrically at 27 °C in 80% dioxane-water mixture (v/v). In all determinations the ionic strength μ was kept constant at 0.1. From the pH- titrant volume data, the acid dissociation constants of the compounds studied were calculated (see Experimental) and the results are summarized in Table 1.

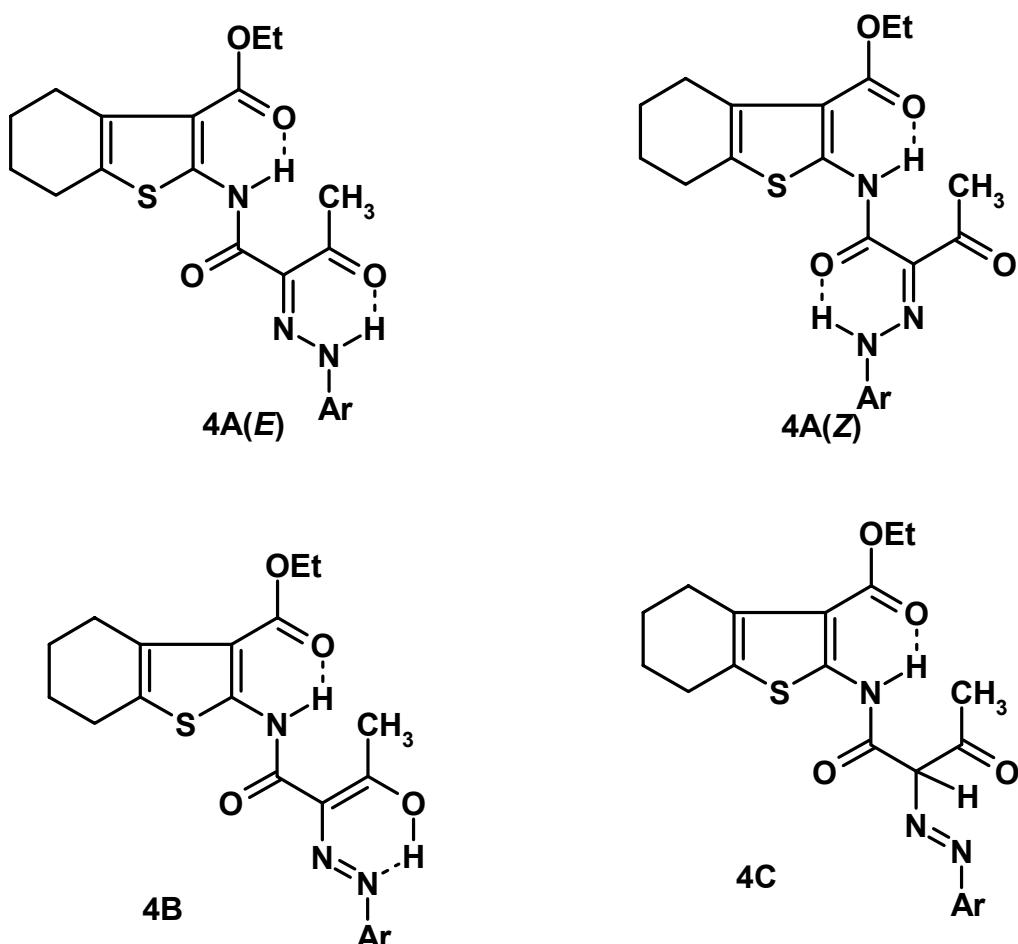


Figure 1. The tautomeric structure of compound **4 A-C**

When the pK_a values were plotted *versus* Hammett substituent constants σ_x^{13} all the substituents fall on the correlation line except the substituents with -R effect, namely the *p*-Cl, *p*-NO₂ and *p*-CH₃CO groups, which are capable of direct interaction with the negatively charged reaction site. However, when the pK_a values were plotted versus σ_x^- constants,¹³ better correlation was obtained. The Hammett type equation of the regression line obtained is:

$$pK_a = 9.05 - 2.30 \sigma_x^- ; r = 0.999; s = \pm 0.04$$

where r is the correlation coefficient and s is the standard deviation.

This excellent linear correlation indicates that the parameter r^- in the Humffray-Ryan equation¹⁴:

$$pK_a = pK_a^0 + \rho \{ \sigma_x + r^- (\sigma_x^- - \sigma_x) \},$$

which gives the contribution of the resonance effect of the substituent varied, is close to unity for the series **4a-i** studied.

The foregoing linear correlation between pK_a values and σ_x^- constants and the values of ρ and r^- found provided further evidence that the studied compounds **4** exist predominantly in the

hydrazone form **4A**. This is because the values of ρ (2.30) and $r^- = 1.00$ are similar to those reported for ionization of phenols ($\rho = 2.67$; $r^- = 1.00$) and anilinium ions ($\rho = 2.77$; $r^- = 1.00$) in 50% ethanol-water mixture.^{15,16} This finding indicates that the negative charge on the anion formed by deprotonation of **4** is largely localized on the N-atom adjacent to the benzene bearing the substituent. This anion will take up a proton on the N-atom bearing the negative charge to give **4A** which is also stabilized by hydrogen bonding. Furthermore, if either form **4B** or **4C** were the predominant form for the studied compounds, the ρ values would be less than 2.0 and will be similar to that reported for ionization of 2-arylazophenols ($\rho = 1.223$; $r^- = 0.286$).¹⁵ Since the bridge between the substituent and the deprotonation reaction site in either form **4B** or **4C** is longer than in **4A**. Thus, it is not unreasonable to conclude that the observed linear correlation of the acid dissociation constants with the Hammett equation indicates that the hydrazone tautomeric form **4A** prevails under the conditions of the pK_a measurement. Compound **4A** can exist in the *E*-structure or the *Z*-structure, the *E*-structure of hydrazone **4A** was preferred than the *Z*-structure¹⁷⁻¹⁹ in which the acetyl carbonyl rather than the amide carbonyl was involved in the H-bonded system.

Table 1. UV Spectral data and acid dissociation constants^a of compounds **4a-i** in dioxane

Compd. no.	λ_{\max} (log ϵ)	pKa ($\pm s$)	σ_x^-
4a	411 (4.76), 255 (4.57)	9.65 (0.02)	-0.27
4b	401 (4.86), 245 (5.30)	9.43 (0.01)	-0.17
4c	396 (4.89), 243 (5.30)	9.20 (0.01)	-0.07
4d^{b)}	396 (4.61), 281 (3.91)	9.08 (0.03)	0.0
4e	400 (4.60), 252 (4.49)	8.55 (0.02)	0.23
4f	394 (4.68), 244 (5.30)	8.18 (0.03)	0.37
4g	395 (4.82), 244 (5.30)	7.43 (0.01)	0.71
4h	411 (4.55), 243 (5.30)	6.10 (0.03)	1.28
4i	395 (4.52), 246 (4.90)	7.57 (0.07)	0.68

^a pK_a in dioxane-water (4:1 v/v) solution at 27°C and $\mu = 0.10$. ^b Solvent λ_{\max} (log ϵ): acetic acid 397 (4.20), 235 (4.65); ethanol 397 (4.78), 244 (4.75); DMF 396 (4.90), 258 (5.30); DMSO 399 (4.87); 251 (5.30).

Pharmacology

Antitumor screening test

The cytotoxic effects of the synthesized compounds **3** and **4** at three different concentrations of each, and doxorubicin (*Adriablastina*), as a reference drug, against *Ehrlich Ascites Carcinoma* tumor cells (EAC) were evaluated in vitro.

As shown in Table 2, the results of antitumor activity screening of the studied compounds indicate that compounds **3**, **4b**, **4e**, **4f** and **4g** have high degree of antitumor activities compared to the positive control toward *Ehrlich Ascites Carcinoma* tumor cells (in vitro). The other derivatives of compounds **4** have moderate or low antitumor activities toward *Ehrlich Ascites Carcinoma* tumor cells (in vitro) in relation with the reference drug *doxorubicin* (Table 2).

Table 2. In vitro cytotoxic activity of compounds **3,4** and doxorubicin against (EAC)

Compound No.	Non-viable cells (%)			
	concentration ($\mu\text{g/ml}$)	100	50	25
3	70	30	10	
4a	27	12	0	
4b	52	20	10	
4c	30	14	7	
4d	36	17	8	
4e	50	24	11	
4f	48	22	10	
4g	70	34	14	
4h	44	21	9	
4i	22	9	0	
Cell control	100%	100%	100%	
doxorubicin	25%	13%	6%	

In conclusion, a simple method for the preparation of compound **3** in 95% yield and their coupling reaction to give new hydrazone derivatives **4a-i** is described. The structures of the newly synthesized compounds **4a-i** were confirmed by spectral and elemental analyses data and the correlation with Hammett equation. These new compounds have been evaluated for the antitumor activity, compounds **3**, **4b**, **4e**, **4f** and **4g** have high antitumor activity against *Ehrlich Ascites Carcinoma* cells. The other derivatives in series **4** exhibit moderate or low activity.

Experimental Section

General Procedures. Melting points were determined on a Gallenkamp apparatus and are uncorrected. IR spectra were recorded in potassium bromide using Perkin Elmer FTIR 1650 and Pye-Unicam SP300 infrared spectrophotometers. ^1H NMR spectra were recorded in deuterated dimethylsulfoxide using a Varian Gemini 300 NMR spectrometer. Mass spectra were recorded on a GCMS-QP 1000 EX Shimadzu and GCMS 5988-A HP spectrometers. Electronic absorption spectra were recorded on Perkin-Elmer Lambda 40 spectrophotometer. Elemental analyses were

carried out at the Microanalytical Laboratory of Cairo University, Giza, Egypt. Antitumor screening was done in the National Institute of Cancer, Cairo, Egypt.

Reaction of ethyl acetoacetate with ethyl 2-amino-4,5,6,7-tetrahydrobenzo [b]thiophene-3-carboxylate (1). Ethyl acetoacetate **2** (2.93 g, 0.023 mole) was heated in conical flask at 100°C, then ethyl 2-amino-4,5,6,7-tetrahydrobenzo[b]thiophene-3-carboxylate **1** (3.38 g, 0.015 mol) was added portionwise with stirring. The temperature was then raised up to 150-160 °C for 20 min. the reaction mixture was left to cool, the solid product was filtered off, washed with ethanol and recrystallized from ethanol to give compound **3** as yellow crystals mp. 80 °C (lit. mp. 90-92 °C in 15 – 20% yield¹⁰); yield 4.40 g (95%). IR ν/cm^{-1} : 3163, 2981, 2939, 2843, 1709, 1682, 1666 cm^{-1} . ^1H NMR (DMSO-d₆): δ/ppm : 1.31 (t, $J = 7.0$ Hz, 3H, CH₃), 1.71 (m, 4H, 2CH₂), 2.20 (s, 3H, COCH₃), 2.59-2.70 (m, 4H, 2CH₂), 3.85 (s, 2H, CH₂), 4.28 (q, $J = 7.0$ Hz, 2H, CH₂), 11.43 (s, 1H, NH). MS m/z (%) 311 ($M^{+}+2$, 8), 310 ($M^{+}+1$, 24), 309 (M^{+} , 57), 225 (64), 179 (100), 151(24), 91 (7). Anal. Calcd. For C₁₅H₁₉NO₄S (309.38) C, 58.23; H, 6.19; N, 4.53%. Found: C, 58.16; H, 6.03; N, 4.32 %.

Preparation of compounds (4a-i)

Method A. To a stirred solution of compound **3** (3.09 g, 10 mmol) in ethanol (50 mL) was added sodium hydroxide (0.4 g, 10 mmol) and the mixture was cooled in an ice bath to 0-5 °C. To the resulting solution, while being stirred, was added dropwise over a period of 20 min., a solution of the appropriate arenediazonium chloride, prepared as usual by diazotizing the respective aniline (10 mmol) in hydrochloric acid (6 M, 6 mL) with sodium nitrite (1M, 10 mL). The whole mixture was then left overnight in a refrigerator. The precipitate solid was collected, washed with water and finally crystallized from the appropriate solvent to give the respective hydrazones (**4a-i**)

Method B. A mixture of **1** (5 mmole) with ethyl 2-arylhydrazone-3-oxobutanoate **5a,d** (5 mmole) in dioxan (30 mL) and triethyl amine (0.35 mL) was heated to reflux for 5 h, then cooled. The solid formed was collected by filtration and crystallized from the appropriated solvent to give the corresponding compounds **4a** and **4d** which was found identical in all respects with that produced by Method A.

N-(3-Ethoxycarbonyl-4,5,6,7-tetrahydrobenzo[b]thien-2-yl)-2-(4-methoxyphenyl hydrazone)-3-oxobutanamide (4a). Orange solid, m.p. 216 °C (Dioxane); yield 3.77 g (85%). IR ν/cm^{-1} : 3409, 3070, 2927, 2835, 1681, 1651, 1629 cm^{-1} . ^1H NMR (DMSO-d₆): δ/ppm : 1.34 (t, $J = 7.0$ Hz, 3H, CH₃), 1.73 (m, 4H, 2CH₂), 2.49 (s, 3H, CH₃), 2.51-2.73 (m, 4H, 2CH₂), 3.78 (s, 3H, CH₃), 4.35 (q, $J = 7.0$ Hz, 2H, CH₂), 7.01 (d, $J = 8.0$ Hz, 2H, Ar-H), 7.54 (d, $J = 8.0$ Hz, 2H, Ar-H), 13.33 (s, 1H, NH), 14.00 (s, 1H, NH). MS m/z (%) 444 ($M^{+}+1$, 17), 443 (M^{+} , 45), 226 (11), 225 (100), 179 (33), 122 (32), 107 (11), 77 (7). Anal. Calcd. For C₂₂H₂₅N₃O₅S (443.52) C, 59.58; H, 5.68; N, 9.47%. Found: C, 59.26; H, 5.80; N, 9.56 %.

N-(3-Ethoxycarbonyl-4,5,6,7-tetrahydrobenzo[b]thien-2-yl)-2-(4-methylphenyl hydrazone)-3-oxobutanamide (4b). Orange crystals, m.p. 202 °C (Dioxane); yield 3.84 g (90%). IR ν/cm^{-1} :

3425, 3078, 2931, 2854, 1682, 1655, 1616 cm^{-1} . ^1H NMR (DMSO-d₆): δ/ppm : 1.34 (t, $J = 7.0$ Hz, 3H, CH₃), 1.73 (m, 4H, 2CH₂), 2.31 (s, 3H, CH₃), 2.49 (s, 3H, CH₃), 2.60-2.73 (m, 4H, 2CH₂), 4.36 (q, $J = 7.0$ Hz, 2H, CH₂), 7.24 (d, $J = 9.0$ Hz, 2H, Ar-H), 7.45 (d, $J = 9.0$ Hz, 2H, Ar-H), 13.32 (s, 1H, NH), 13.98 (s, 1H, NH). MS m/z (%) 428 (M⁺+1, 21), 427 (M⁺, 49), 225 (100), 179 (51), 151 (15), 106 (32), 91 (25), 77 (19); Anal. Calcd. For C₂₂H₂₅N₃O₄S (427.52) C, 61.81; H, 5.89; N, 9.83%. Found: C, 61.74; H, 5.66; N, 9.78 %.

N-(3-Ethoxycarbonyl-4,5,6,7-tetrahydrobenzo[b]thien-2-yl)-2-(3-methylphenyl hydrazone)-3-oxobutanamide (4c). Orange solid, m.p. 180 °C (Dioxane); yield 3.50 g (82%). IR ν/cm^{-1} : 3454, 3129, 2941, 2861, 1682, 1666, 1655 cm^{-1} . ^1H NMR (DMSO-d₆): δ/ppm : 1.31 (t, $J = 7.0$ Hz, 3H, CH₃), 1.70 (m, 4H, 2CH₂), 2.33 (s, 3H, CH₃), 2.50 (s, 3H, CH₃), 2.51-2.68 (m, 4H, 2CH₂), 4.33 (q, $J = 7.0$ Hz, 2H, CH₂), 7.00-7.30 (m, 4H, Ar-H), 13.26 (s, 1H, NH), 13.93 (s, 1H, NH). MS m/z (%) 428 (M⁺+1, 31), 427 (M⁺, 45), 225 (100), 178 (16), 106 (23), 91 (26), 77 (17). Anal. Calcd. For C₂₂H₂₅N₃O₄S (427.52) C, 61.81; H, 5.89; N, 9.83%. Found: C, 61.63; H, 5.44; N, 9.58 %.

N-(3-Ethoxycarbonyl-4,5,6,7-tetrahydrobenzo[b]thien-2-yl)-2-phenylhydrazone-3-oxobutanamide (4d). Orange crystals, m.p. 208 °C (Dioxane/Ethanol); yield 3.80 g (92%). IR ν/cm^{-1} : 3429, 3074, 2928, 2851, 1681, 1659, 1624 cm^{-1} . ^1H NMR (DMSO-d₆): δ/ppm : 1.34 (t, $J = 7.0$ Hz, 3H, CH₃), 1.72 (m, 4H, 2CH₂), 2.49 (s, 3H, CH₃), 2.60-2.72 (m, 4H, 2CH₂), 4.35 (q, $J = 7.0$ Hz, 2H, CH₂), 7.21-7.57 (m, 5H, Ar-H), 13.30 (s, 1H, NH), 13.94 (s, 1H, NH). MS m/z (%) 414 (M⁺+1, 50), 413 (M⁺, 64), 225 (100), 224 (12), 179 (66), 151 (15), 92 (24), 91 (11), 77 (22). Anal. Calcd. For C₂₁H₂₃N₃O₄S (413.49) C, 61.00; H, 5.61; N, 10.16%. Found: C, 60.98; H, 5.40; N, 10.00 %.

N-(3-Ethoxycarbonyl-4,5,6,7-tetrahydrobenzo[b]thien-2-yl)-2-(4-chlorophenyl hydrazone)-3-oxobutanamide (4e). Orange solid, m.p. 214-218 °C (Dioxane); yield 3.80 g (89%). IR ν/cm^{-1} : 3422, 3074, 2928, 2850, 1681, 1658, 1630 cm^{-1} . ^1H NMR (DMSO-d₆): δ/ppm : 1.34 (t, $J = 7.0$ Hz, 3H, CH₃), 1.72 (m, 4H, 2CH₂), 2.50 (s, 3H, CH₃), 2.61 (m, 2H, CH₂), 2.73 (m, 2H, CH₂), 4.36 (q, $J = 7.0$ Hz, 2H, CH₂), 7.47 (d, $J = 9.0$ Hz, 2H, Ar-H), 7.60 (d, $J = 9.0$ Hz, 2H, Ar-H), 13.30 (s, 1H, NH), 13.88 (s, 1H, NH). MS m/z (%) 449 (M⁺+2, 30), 447 (M⁺, 40), 226 (20), 225 (100), 179 (53), 178 (14), 126 (17), 111 (13), 91 (16), 77 (9); Anal. Calcd. For C₂₁H₂₂ClN₃O₄S (447.94): C, 56.31; H, 4.95; N, 9.38%. Found: C, 56.30; H, 4.86; N, 9.24 %.

N-(3-Ethoxycarbonyl-4,5,6,7-tetrahydrobenzo[b]thien-2-yl)-2-(3-chlorophenyl hydrazone)-3-oxobutanamide (4f). Orange solid, m.p. 205 °C (Dioxane); yield 3.53 g (79%). IR ν/cm^{-1} : 3429, 3105, 2977, 2931, 2854, 1680, 1654, 1624 cm^{-1} . ^1H NMR (DMSO-d₆): δ/ppm : 1.34 (t, $J = 7.0$ Hz, 3H, CH₃), 1.70 (m, 4H, 2CH₂), 2.48 (s, 3H, CH₃), 2.51-2.69 (m, 4H, 2CH₂), 4.34 (q, $J = 7.0$ Hz, 2H, CH₂), 7.20-7.59 (m, 4H, Ar-H), 13.24 (s, 1H, NH), 13.79 (s, 1H, NH). MS m/z (%) 449 (M⁺+2, 20), 448 (M⁺+1, 15), 447 (M⁺, 61), 226 (24), 225 (100), 180 (13), 179 (67), 151 (17), 126 (22), 111 (12), 91 (8). Anal. Calcd. For C₂₁H₂₂ClN₃O₄S (447.94): C, 56.31; H, 4.95; N, 9.38%. Found: C, 56.20; H, 4.71; N, 9.09 %.

N-(3-Ethoxycarbonyl-4,5,6,7-tetrahydrobenzo[b]thien-2-yl)-2-(3-nitrophenyl hydrazone)-3-oxobutanamide (4g). Dark yellow solid, m.p. 220 °C (Dioxane); yield 3.57 g (78%). IR ν/cm^{-1} :

3409, 3066, 2981, 2927, 2881, 1680, 1662, 1624 cm^{-1} . ^1H NMR (DMSO-d₆): δ/ppm : 1.34 (t, J = 7.0 Hz, 3H, CH₃), 1.70 (m, 4H, 2CH₂), 2.49 (s, 3H, CH₃), 2.57-2.66 (m, 4H, 2CH₂), 4.33 (q, J = 7.0 Hz, 2H, CH₂), 7.65-7.832 (m, 4H, Ar-H), 13.19 (s, 1H, NH), 13.82 (s, 1H, NH). MS m/z (%) 458 (M⁺, 100), 226 (14), 225 (77), 179 (53), 151 (25), 127 (14), 125 (28), 91 (11), 77 (10). Anal. Calcd. For C₂₁H₂₂N₄O₆S (458.49): C, 55.01; H, 4.84; N, 12.22%. Found: C, 54.91; H, 4.60; N, 12.06 %.

N-(3-Ethoxycarbonyl-4,5,6,7-tetrahydrobenzo[b]thien-2-yl)-2-(4-nitrophenyl hydrazone)-3-oxobutanamide (4h). Orange solid, m.p. 240 °C (DMF); yield 4.08 g (89%). IR ν/cm^{-1} : 3406, 3058, 2981, 2931, 2835, 1701, 1670, 1654, cm^{-1} . ^1H NMR (DMSO-d₆): δ/ppm : 1.34 (t, J = 7.0 Hz, 3H, CH₃), 1.73 (m, 4H, 2CH₂), 2.49 (s, 3H, CH₃), 2.54-2.72 (m, 4H, 2CH₂), 4.35 (q, J = 7.0 Hz, 2H, CH₂), 7.73 (d, J = 9.0 Hz, 2H, Ar-H), 8.24 (d, J = 9.0 Hz, 2H, Ar-H), 13.23 (s, 1H, NH), 13.87 (s, 1H, NH). MS m/z (%) 459 (M⁺+1, 24), 458 (M⁺, 60), 225 (100), 206 (18), 180 (23), 179 (69), 178 (20), 151 (22), 150 (18), 137 (11), 77 (11). Anal. Calcd. For C₂₁H₂₂N₄O₆S (458.49): C, 55.01; H, 4.84; N, 12.22%. Found: C, 54.93; H, 4.59; N, 12.15 %.

N-(3-Ethoxycarbonyl-4,5,6,7-tetrahydrobenzo[b]thien-2-yl)-2-(4-acetylphenyl hydrazone)-3-oxobutanamide (4i). Red solid, m.p. 236-237 °C (Dioxane); yield 3.91 g (89%). IR ν/cm^{-1} : 3448, 3087, 2981, 2939, 2864, 1682, 1671, 1635, 1624 cm^{-1} . ^1H NMR (DMSO-d₆): δ/ppm : 1.33 (t, J = 7.0 Hz, 3H, CH₃), 1.75 (m, 4H, 2CH₂), 2.49 (s, 3H, CH₃), 2.56 (s, 3H, CH₃), 2.65-2.76 (m, 4H, 2CH₂), 4.38 (q, J = 7.0 Hz, 2H, CH₂), 7.80 (d, J = 9.0 Hz, 2H, Ar-H), 8.28 (d, J = 9.0 Hz, 2H, Ar-H), 13.29 (s, 1H, NH), 13.89 (s, 1H, NH). MS m/z (%) 456 (M⁺+1, 6), 455 (M⁺, 60), 321 (25), 245 (16), 226 (16), 204 (18), 179 (16), 178 (18), 151 (14), 150 (11), 120 (48), 119 (86), 106 (33), 91 (100), 77 (43). Anal. Calcd. For C₂₃H₂₅N₃O₅S (455.53): C, 60.64; H, 5.53; N, 9.22%. Found: C, 60.45; H, 5.41; N, 9.05 %.

pK_a Determination of Compounds (4a-i)

The acid dissociation constants of compounds **4** were determined potentiometrically in 80% dioxan-water mixture at 27±0.1 °C and ionic strength (KNO₃) μ 0.1. A Metrohm 686 titroprocessor equipped with 665 Dosimat was used. The electrode and the titroprocessor were calibrated with standard Beckman buffer solutions of pH 4.01 and 7.00. The pH meter reading B recorded in dioxane-water solution was converted to hydrogen ion concentration [H⁺] by means of the relation of van Uitert and Hass²⁰ namely:

$$-\log [\text{H}^+] = \text{B} + \log \text{U}_\text{H}$$

Where log U_H is the correction factor for the solvent composition and ionic strength used for which B is read. The value of log U_H was found to be 0.48. A carbonate-free sodium hydroxide titrant was prepared and standardized against potassium hydrogen phthalate solution.

The experimental procedure followed in the determination of *pK_a* values and their calculations, by the method of least squares, from the titrant volume-pH data using the relation: $pK_a = \text{pH}_i - \log V_i / (V_e - V_i)$, is similar to that previously described.²¹ In this equation, pH_i is the corrected pH value of the solution when the volume of the added titrant is V_i and V_e is the volume of the titrant at the equivalence point. The calculations of the *pK_a* values were carried out using

computer program MINIQUAD-75.²² The pK_a values obtained were reproducible to within ± 0.02 pK_a unit. The results are summarized in Table 1.

Cytotoxic activity against *Ehrlich Ascites Carcinoma tumor cell (EAC)* in vitro²³: *Ehrlich Ascites Carcinoma cell* (EAC) used in this study was kindly supplied by National Institute of Cancer in Netherland since 1952 and maintained in Swiss albino mice by serial I P transplantation , RPMI 1640 medium (Sigma), *Ehrlich Ascites Carcinoma cell* (EAC) suspension (2.5×10^5 cells/ml), and Trypan blue dye were used. A stock solution was prepared by dissolving one gram of the dye in distilled water (100 ml). The working solution was then prepared by diluting (1 ml) of the stock solution with 9 ml of distilled water. The stain was then used for staining the dead EAC cells.

A set of sterile test tubes was used, where 2.5×10^5 tumor cells / ml were suspended in phosphate buffered saline (PBS). Then 25, 50 and 100 μg / ml from tested compound were added to the suspension, kept at 37 °C for 2 h. Trypan blue dye exclusion test was then carried out to calculate the percentage of nonviable cells. Doxorubicin (Adriablastina) was used as a positive control. The percentage of the nonviable cells was calculated by the following equation:

% of non-viable cells = 100 (N / N_t), where N is the number of non-viable cells counted, N_t is the total number of cells. The test was repeated four times for each compound. The results are summarized in Table 2.

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